CELL WALL SYNTHESIS IN STAPHYLOCOCCUS AUREUS IN THE PRESENCE OF PROTEIN SYNTHESIS INHIBITORY AGENTS

TETRACYCLINE AND AMINOGLYCOSIDE ANTIBIOTICS

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Abstract

Morphological changes of *Staphylococcus aureus*, exposed to protein synthesis inhibitory antibiotics such as tetracyclines and aminoglycosides were studied by electron-microscopic observation using ultra-thin sections. Cell wall thickening of *S. aureus* was observed by these antibiotics, and abnormal cell division was remarkably noticed with tetracyclines. With clinically isolated *S. aureus*, resistant to each antibiotic, the phenomena of cell wall thickening and abnormal cell division were not observed when exposed to that antibiotic.

In previous publication^{1,2,3)}, we reported that the cell wall of *S. aureus* became so thickened and often accompanied with the formation of multilayers, and the electron-density of cytoplasm was increased following exposure to protein synthesis inhibitory antibiotics such as lincomycins and macrolides, and these phenomena were not observed with clinically isolated resistant strains. On the other hand, this phenomenon of cell wall thickening was also confirmed in biochemical study⁴⁾. This time we investigated electron-microscopically the morphological changes of *S. aureus* treated with tetracycline and aminoglycoside antibiotics.

Materials and Methods

As the test organism, Staphylococcus aureus 209-P JC and clinically isolated three strains of Staphylococcus aureus; (S. aureus No. 8, resistant to tetracycline, S. aureus No. 12, resistant to streptomycin, S. aureus No. 7, resistant to both streptomycin and kanamycin) were used. Cells were preincubated in Tryptosoya Broth (TSB, Nissan) for 18 hours, and diluted by Heart Infusion Broth (HIB, Nissan), and mixed with Heart Infusion Agar (HIA, Nissan) to make plates containing about 10⁶ cells/ml. Cylinders were placed on the plates, then tetra-

cycline (TC; Lederle (Japan) Ltd.), minocycline (MINO, Lederle (Japan) Ltd.), streptomycin (SM, Meiji Seika Kaisha, Ltd.) and kanamycin (KM, Meiji Seika Kaisha, Ltd.), (10 mcg/ml respectively) were poured into each cylinder. The plates were cultivated for about 24 hours at 37°C to make the inhibitory circle, and the boundary of the circle was cut out to obtain agar blocks. With resistant strains the inhibitory circle was not produced, so the agar just outside the cylinder was cut out. Agar blocks were fixed with 1% OsO4 dissolved in KELLENBERGER buffer⁵⁾ at room temperature for 18 hours. After the fixation, blocks were stained with 0.5% of uranyl acetate and dehydrated by alcohol series, then embedded in epoxy resin⁶). Ultra-thin sections, obtained by ultra-microtome (LKB), were stained with uranyl acetate and lead citrate⁷⁾, then observed by electron microscope AKASHI S-500.

Results and Discussion

Fig. 1 shows the electron-micrograph of intact S. aureus 209-P JC. Following exposure to tetracycline antibiotics the phenomena of cell wall thickening, abnormal cell division and the increase of electron-density in cytoplasm were observed (Fig. 2, 3). With aminoglycoside antibiotics such as SM, KM the phenomenon of peripheral cell wall thickening was also observed, but abnormal cell division or the increase of electron-density in cytoplasm were not noticed (Fig. 4, 5). In the case of clinically isolated strain which shows resistance to TC, morphological changes such as in sensitive strain were not noticed with following exposure to TC (Fig. 6), but typical phenomena with tetracycline antibiotics were noticed when treated with MINO (Fig. 7). The relation between the cross resistance in aminoglycoside antibiotics and the phenomenon of cell wall thickening was also the same in the case of tetracyclines (Fig. 8, 9).

The results obtained in this experiment are



Fig. 1. Intact S. aureus 209-P JC. ×119,000 Nuclear division is finished and cross walls growing are observed.



Fig. 2. S. aureus 209-P JC, treated with TC. $\times 114,000$ Cell walls are thickened and normal binary division is disrupted. Electron-density in cytoplasm is increased.



Fig. 3. S. aureus 209-P JC, treated with MINO. $\times 117,000$ Peripheral cell wall is thickened and layers are seen in it. Electron-density in cytoplasm is increased.



Fig. 4. S. aureus 209-P JC, treated with SM. $\times 131,000$ Peripheral cell wall is thickened but the thickness of cross wall is not so changed compared with intact cell.



Fig. 5. S. aureus 209-P JC, treated with KM. $\times 120,000$ The degree of cell wall thickening is about the same as in the case of SM.



Fig. 6. S. aureus No. 39, treated with TC. $\times 139,000$ Morphological changes observed in sensitive strain are not seen.



Fig. 7. S. aureus No. 39, treated with MINO. $\times 107,000$

The phenomena of cell wall thickening, abnormal cell division and the increase of electron-density in cytoplasm which are characteristic of tetracycline antibiotics are observed.



Fig. 9. S. aureus No. 12, treated with KM. $\times 131,000$ Peripheral cell wall is thickened and mesosomes in contact with the slightly thickened cross wall of growing state are seen.

summarized as follows: cell wall of *S. aureus* became thickened following exposure to tetracycline and aminoglycoside antibiotics. With tetracylines, abnormal cell division and increased electron-density in cytoplasm were not noticed. In this study, the degree of cell wall thickening of cells treated with tetracyclines or aminoglycosides was not so extreme compared with the cells treated with macrolide antibiotics. Relation between the cross resistance of drugs and the phenomenon of cell wall thickening was the same as in the case of macrolides.

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Fig. 8. S. aureus No. 12, treated with SM. $\times 117,000$ Cell wall thickening is not seen.

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References

- NAKAO, M.; E. KITANAKA, K. OCHIAI & S. NAKAZAWA: Cell wall synthesis by Staphylococcus aureus in the presence of protein synthesis inhibitory agents. I. Electron microscopic study. J. Antibiotics 25: 60~63, 1972
- NAKAO, M.; E. KITANAKA, K. OCHIAI & S. NAKAZAWA: Cell wall synthesis by Staphylococcus aureus in the presence of protein synthesis inhibitory agents. II. Electron microscopic study. J. Antibiotics 25: 469~470, 1972
- NAKAO, M.; E. KITANAKA, K. OCHIAI & S. NAKAZAWA: Cell wall synthesis in Staphylococcus aureus in the presence of protein synthesis inhibitory agents. I. Lincomycin, clindamycin and macrolide antibiotics. Japan. J. Microbiol. 16: 403~413, 1972
- 4) KITANAKA, E.; K. OCHIAI, M. NAKAO & S. NAKAZAWA: Cell wall synthesis by Staphylococcus aureus in the presence of protein synthesis inhibitory agents. III. Biochemical study. J. Antibiotics 25: 679~680, 1972
- 5) KELLENBERGER, E.; A. RYTER & J. SECHAUD: Electron microscope study of DNA-containing plasms. II. Vegetative and mature phage DNA as compared with normal bacterial nucleotide in different physiological states. J. Biophys. Biochem. Cytol. 4:671~678, 1958
- 6) LUFT, J. H.: Improvements in epoxy resin embedding methods. J. Biophys. Biochem. Cytol. 9:409~414, 1961
- REYNOLDS, E. S.: The use of lead citrate at high pH as an electron-opaque stain in electron microscopy. J. Cell Biol. 17: 208~212, 1963